

# Evidence for Protein–Polysaccharide Complex Formation as a Result of Dry-heating of Mixtures

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## 1 Introduction

The complexes produced by the dry-heating of the globular protein bovine serum albumin (BSA) and the non-ionic polysaccharide dextran have been shown recently to have excellent emulsion stabilizing properties.<sup>1,2</sup> This short paper presents experimental data on the molecular weight of complexes formed by dry-heating mixtures of BSA and dextran T40 ( $4 \times 10^4$  daltons) as determined by gel permeation chromatography with light-scattering detection (GPC/MALLS) and ultracentrifugation.

## 2 Experimental

Protein–polysaccharide complexes were prepared by dry heating various molar ratios of BSA and dextran T40 at 60 °C for 3 weeks as described previously.<sup>1</sup> The samples were dissolved in phosphate/chloride buffer (pH 7.0, ionic strength 0.1 M). Weight-average molecular weights of samples of various molar ratios were determined by GPC/MALLS (sample concentration 3 mg ml<sup>-1</sup>).<sup>3</sup> The BSA–dextran complex of molar ratio 2:1 in a solution of concentration 0.5 mg ml<sup>-1</sup> was investigated under conditions of low-speed sedimentation equilibrium in a Model E analytical ultracentrifuge. The sedimentation coefficient of this same complex (sample concentration 2 mg ml<sup>-1</sup>) was determined using a Beckman XL-A analytical ultracentrifuge.

### 3 Results and Discussion

Figure 1 shows the weight-average molecular weight  $M_w$  of various BSA-dextran complexes as determined by GPC/MALLS. We see that, of the various samples investigated, only the complex with a composition of 33 mol% dextran has a molecular weight substantially greater than that for the native protein. Table 1 compares the value of  $M_w$  obtained from GPC/MALLS for this 2:1 molar ratio complex with that from low-speed sedimentation equilibrium. Also shown for comparison are  $M_w$  values for the heat-treated pure BSA (same heating conditions). The results indicate that, although some aggregation of the BSA itself occurs during the

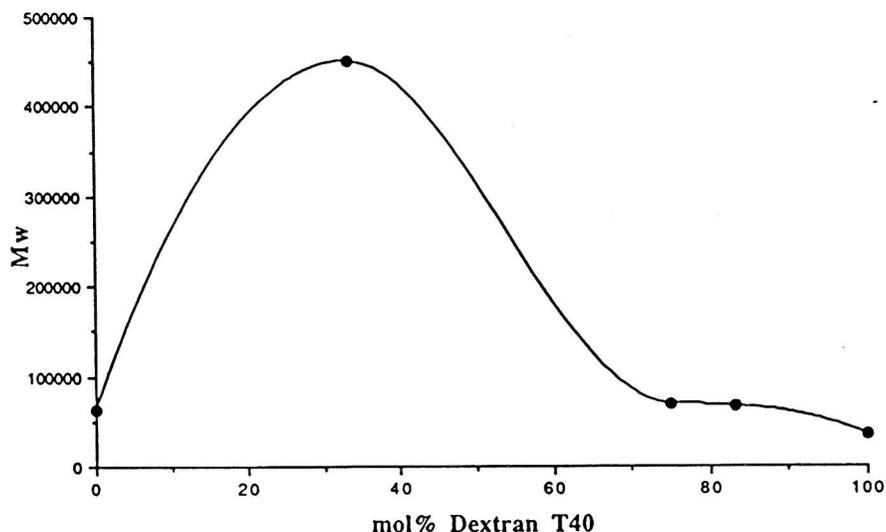


Figure 1 Weight-average molecular weight  $M_w$  of complexes obtained by dry-heating of mixtures of BSA and dextran T40 at various molar ratios

Table 1 Molecular weight, sedimentation coefficients, and frictional coefficients

Sample	$M_w$ (daltons) GPC/MALLS	$M_w$ (daltons) Model E	$s_{20}$ (S)	$s_{20,w}$ (S)	$f/f_0$
BSA native	62 700 ± 5 000	66 700 <sup>a</sup>	3.93 ± 0.1	4.04 ± 0.1	1.3 ± 0.1
BSA heat-treated (3 weeks)	158 000 ± 10 000	130 000 ± 10 000	5.43 ± 0.1	5.59 ± 0.1	1.3 ± 0.1
BSA/T40 complex 2:1 ratio	450 000 ± 20 000	330 000 ± 20 000	4.85 ± 0.1	4.98 ± 0.1	3.4 ± 0.1

<sup>a</sup> Value from: K. E. van Holde, 'Physical Biochemistry', Prentice-Hall, 1971

dry-heating process, this does not account for the much larger average molecular weight of the complex. The general trends of behaviour detected by GPC/MALLS and ultracentrifugation are the same. The slight discrepancy in the numerical values can be accounted for by noting that the light-scattering results may be affected to some extent by additional aggregated material co-existing with the protein-polysaccharide complex, whereas such material might have moved to the cell base in ultracentrifugation measurements and would therefore not have contributed to the weight-average molecular weight.

Figure 2 compares chromatographic elution profiles for the BSA-dextran complex (2:1 molar ratio) with those for the native BSA and the heat-treated BSA. We note the different position of the complex peak (lower elution volume) from that for the protein alone, both from light-scattering analysis and differential refractive index analysis. The peak shape shows that the complex is not a single species but is polydisperse. Figure 3 shows the movement of the sedimenting boundary for the BSA-dextran complex (2:1 molar ratio). The presence of a single sedimenting boundary suggests that the polydispersity is more of a quasi-continuous character than a paucidisperse one.

Sedimentation coefficients of complex, native BSA and heated BSA are also given in Table 1. From the molecular weight and the  $s_{20,w}$  value, we can estimate the frictional ratio  $f/f_0$ . Assuming that the  $s_{20,w}$  value measured is not too far from the infinite dilution value, the large value of  $f/f_0$  derived for the complex is suggestive of either a highly asymmetric or a highly hydrated entity.

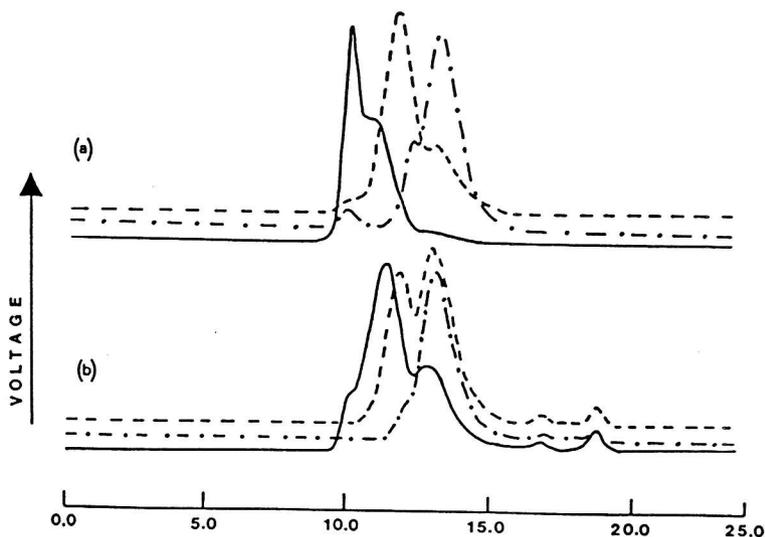


Figure 2 (a) Light-scattering chromatograms (obtained from  $90^\circ$  detector) and (b) Differential refractive index chromatograms:  $\cdots$ , native BSA;  $---$ , heat treated BSA;  $-$ , BSA/dextran T40 (2:1) complex

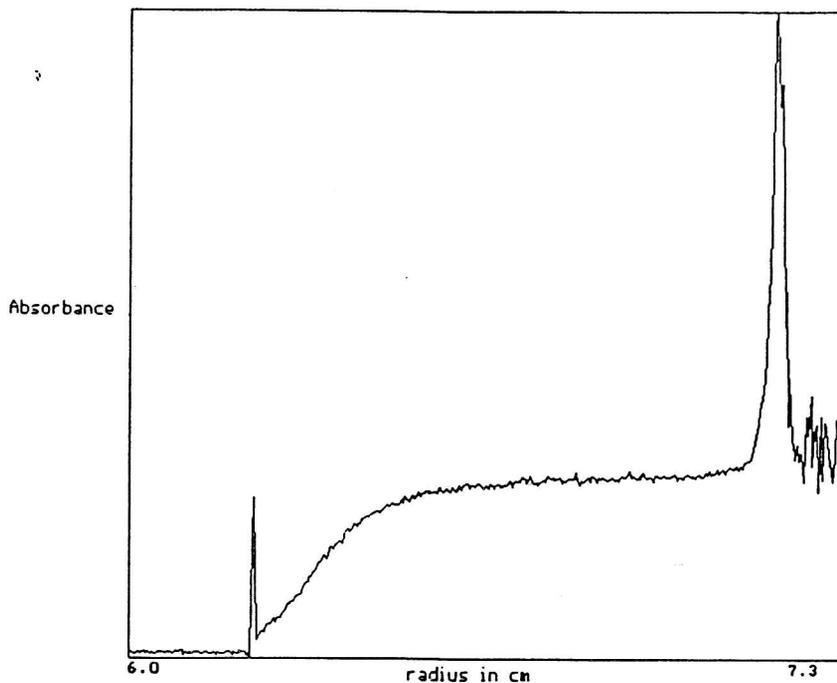


Figure 3 *Sedimenting boundary of BSA/dextran T40 (2:1) complex*

## References

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