

Evidence for Protein–Polysaccharide Complex Formation as a Result of Dry-heating of Mixtures

By K. Jumel, S. E. Harding, J. R. Mitchell, and Eric Dickinson¹

DEPARTMENT OF APPLIED BIOCHEMISTRY AND FOOD SCIENCE, UNIVERSITY OF NOTTINGHAM, SUTTON BONINGTON, LOUGHBOROUGH, LEICESTERSHIRE LE12 5RD, UK

¹PROCTER DEPARTMENT OF FOOD SCIENCE, UNIVERSITY OF LEEDS, LEEDS LS2 9JT, UK

1 Introduction

The complexes produced by the dry-heating of the globular protein bovine serum albumin (BSA) and the non-ionic polysaccharide dextran have been shown recently to have excellent emulsion stabilizing properties.^{1,2} This short paper presents experimental data on the molecular weight of complexes formed by dry-heating mixtures of BSA and dextran T40 (4×10^4 daltons) as determined by gel permeation chromatography with light-scattering detection (GPC/MALLS) and ultracentrifugation.

2 Experimental

Protein–polysaccharide complexes were prepared by dry heating various molar ratios of BSA and dextran T40 at 60 °C for 3 weeks as described previously.¹ The samples were dissolved in phosphate/chloride buffer (pH 7.0, ionic strength 0.1 M). Weight-average molecular weights of samples of various molar ratios were determined by GPC/MALLS (sample concentration 3 mg ml⁻¹).³ The BSA–dextran complex of molar ratio 2:1 in a solution of concentration 0.5 mg ml⁻¹ was investigated under conditions of low-speed sedimentation equilibrium in a Model E analytical ultracentrifuge. The sedimentation coefficient of this same complex (sample concentration 2 mg ml⁻¹) was determined using a Beckman XL-A analytical ultracentrifuge.

3 Results and Discussion

Figure 1 shows the weight-average molecular weight M_w of various BSA-dextran complexes as determined by GPC/MALLS. We see that, of the various samples investigated, only the complex with a composition of 33 mol% dextran has a molecular weight substantially greater than that for the native protein. Table 1 compares the value of M_w obtained from GPC/MALLS for this 2:1 molar ratio complex with that from low-speed sedimentation equilibrium. Also shown for comparison are M_w values for the heat-treated pure BSA (same heating conditions). The results indicate that, although some aggregation of the BSA itself occurs during the

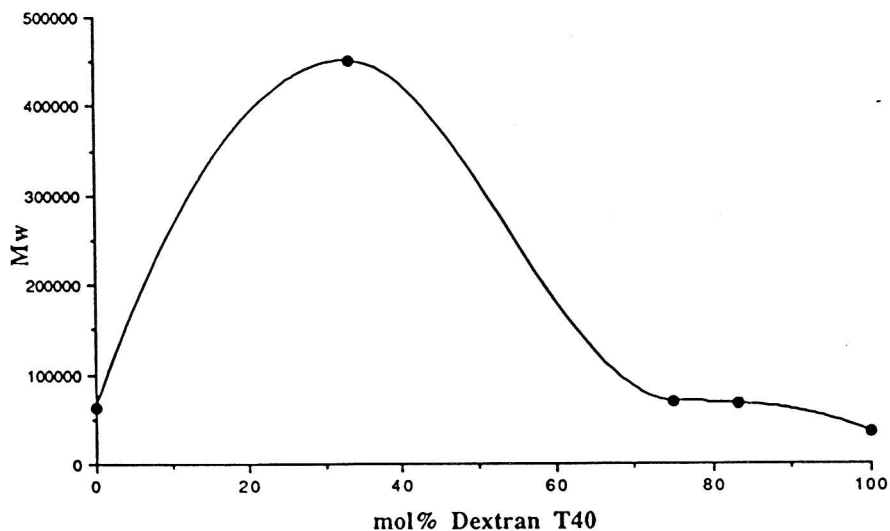


Figure 1 Weight-average molecular weight M_w of complexes obtained by dry-heating of mixtures of BSA and dextran T40 at various molar ratios

Table 1 Molecular weight, sedimentation coefficients, and frictional coefficients

Sample	M_w (daltons) GPC/MALLS	M_w (daltons) Model E	s_{20} (S)	$s_{20,w}$ (S)	f/f_0
BSA native	62 700 ± 5 000	66 700 ^a	3.93 ± 0.1	4.04 ± 0.1	1.3 ± 0.1
BSA heat-treated (3 weeks)	158 000 ± 10 000	130 000 ± 10 000	5.43 ± 0.1	5.59 ± 0.1	1.3 ± 0.1
BSA/T40 complex 2:1 ratio	450 000 ± 20 000	330 000 ± 20 000	4.85 ± 0.1	4.98 ± 0.1	3.4 ± 0.1

^a Value from: K. E. van Holde, 'Physical Biochemistry', Prentice-Hall, 1971

dry-heating process, this does not account for the much larger average molecular weight of the complex. The general trends of behaviour detected by GPC/MALLS and ultracentrifugation are the same. The slight discrepancy in the numerical values can be accounted for by noting that the light-scattering results may be affected to some extent by additional aggregated material co-existing with the protein-polysaccharide complex, whereas such material might have moved to the cell base in ultracentrifugation measurements and would therefore not have contributed to the weight-average molecular weight.

Figure 2 compares chromatographic elution profiles for the BSA-dextran complex (2:1 molar ratio) with those for the native BSA and the heat-treated BSA. We note the different position of the complex peak (lower elution volume) from that for the protein alone, both from light-scattering analysis and differential refractive index analysis. The peak shape shows that the complex is not a single species but is polydisperse. Figure 3 shows the movement of the sedimenting boundary for the BSA-dextran complex (2:1 molar ratio). The presence of a single sedimenting boundary suggests that the polydispersity is more of a quasi-continuous character than a paucidisperse one.

Sedimentation coefficients of complex, native BSA and heated BSA are also given in Table 1. From the molecular weight and the $s_{20,w}$ value, we can estimate the frictional ratio f/f_0 . Assuming that the $s_{20,w}$ value measured is not too far from the infinite dilution value, the large value of f/f_0 derived for the complex is suggestive of either a highly asymmetric or a highly hydrated entity.

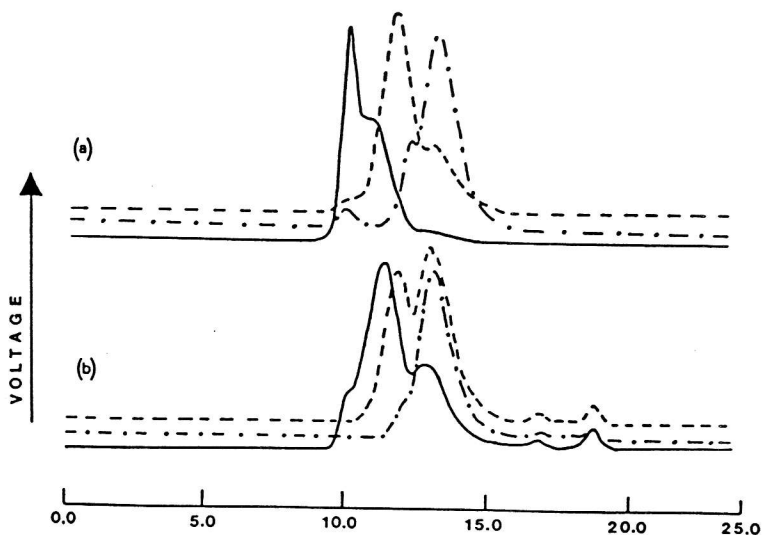


Figure 2 (a) Light-scattering chromatograms (obtained from 90° detector) and (b) Differential refractive index chromatograms: \cdots , native BSA; $---$, heat treated BSA; $-$, BSA/dextran T40 (2:1) complex

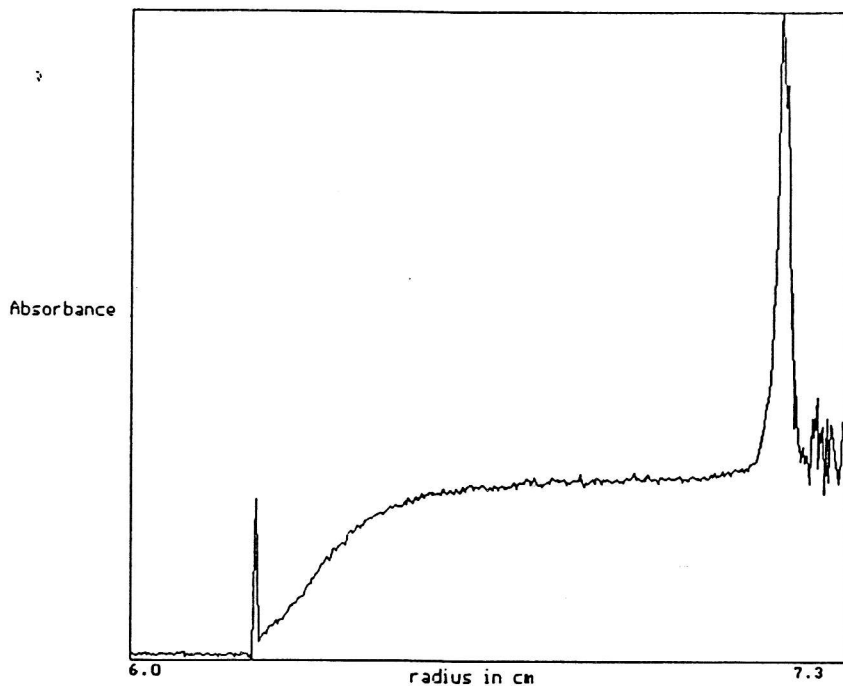


Figure 3 *Sedimenting boundary of BSA/dextran T40 (2:1) complex*

References

1. E. Dickinson and V. B. Galazka, in 'Gums and Stabilisers in the Food Industry', ed. G. O. Phillips, D. J. Wedlock, and P. A. Williams, Oxford University Press, 1992, Vol. 6, p.351.
2. E. Dickinson and M. G. Semenova, *Colloids Surf.*, 1992, **64**, 299.
3. P. J. Wyatt, in 'Laser Light Scattering in Biochemistry', ed. S. E. Harding, D. B. Satelle, and V. A. Bloomfield, Special Publication No. 99, Royal Society of Chemistry, Cambridge, 1992, p. 35.